

Journal of Coastal Life Medicine

journal homepage: www.jclmm.com



Review doi: 10.12980/JCLM.3.2015JCLM-2015-0018

©2015 by the Journal of Coastal Life Medicine. All rights reserved.

An overview of the marine natural products in clinical trials and on the market

Marisa Rangel^{1,2*}, Miriam Falkenberg³

¹Immunopathology Laboratory, Division of Scientific Development, Butantan Institute, 05503-900, Sao Paulo, Brazil

²Laboratory of Toxinology, Department of Physiological Sciences, University of Brasilia, 70910-900, Brasilia, DF, Brazil

³Department of Pharmaceutical Sciences, Federal University of Santa Catarina, 88040-970, Florianopolis, SC, Brazil

ARTICLE INFO

Article history:

Received 17 Mar 2015

Accepted 15 Apr 2015

Available online 7 May 2015

Keywords:

Marine natural products

Marine drugs

Anticancer drugs

Antiviral drugs

Analgesic drugs

Clinical trials

ABSTRACT

The first marine natural products that served as leads or scaffolds for medicines were discovered in the middle of last century: the arabinosyl glycosides from the marine sponge *Tectitethya crypta*. Synthesis and modifications of the natural molecules generated antiviral and antileukemic drugs developed in the 1970's and in the following decades, including the first effective treatment against HIV infection.

With the improvement of techniques for the elucidation of chemical structure of the molecules, as well as chemical synthesis, especially from the 1990's, there was an increase in the number of bioactive natural products characterized from marine organisms. New chemical structures with high specificity towards molecular targets in cells allowed the development of new drugs with indication for the treatment of several illnesses, from cancer to new antibiotics, and even neurological disorders.

Currently there are at least 13 molecules derived from marine natural products on advanced clinical trials, and nine were approved to be used as medicines. Considering that in the past eight years, more than 1000 new compounds from marine organisms were described, per year, the expectation is that many more drugs will be derived from marine natural products in a near future.

1. Introduction

The search for biologically active marine natural products started along with the beginning of marine natural products chemistry, with the pioneer work of the researchers Bergmann and Feeney in 1951[1]. When studying a Caribbean marine sponge, *Tectitethya crypta* (*Cryptotethya crypta*), they characterized two arabinosyl glycosides, named spongouridine and spongotimidine (or Ara-U and Ara-T, respectively), with bioactive properties. Once knowing that the biological systems would recognize the nucleoside base with altered sugar moieties, the chemists tried to substitute the typical pentoses by modified sugar or by acyclic entities, leading to the synthesis of several nucleoside analogues used as antitumoral or antiviral drugs, like zidovudine[2], the main antiretroviral drug used to suppress viremia in HIV-positive patients. Obtained by synthesis,

cytarabine (arabinofuranosylcytidine or Ara-C, Cytosar-U®) inserts into the DNA chain, inhibiting the DNA polymerase and holding the cell cycle on phase S; it is indicated for the treatment of acute non-lymphocytic, chronic myelocytic and meningeal leukemias[3,4]. Vidarabine (arabinofuranosyladenine or Ara-A) produced by *Streptomyces antibioticus* fermentation and commercialized as Vira-A®[5], aciclovir (Zovirax®) and other related compounds are indicated for the treatment of herpes simplex and herpes zoster viruses infections[6,7]. Nowadays not only sponges but also other marine organisms (including fungi and bacteria) are being the source of natural compounds with amazing different structures and a myriad of potential activities[8].

The first marine natural product with industrial use was nereistoxin, from the polychaeta worm *Lumbrineris* sp., and used as a prototype in the synthesis of the insecticide Cartap (Padan®)[9], developed by Takeda Chemical Ltd. (Tokyo), and exported to many countries to control insect plagues in orange plantation and other cultures such as rice and sugarcane fields[10,11]. Cartap is considered to be a highly effective, low toxicity and low residue pesticide, with very rare cases of human toxicity resulting from occupational exposure or deliberate self-harm ingestion[11,12].

Like nereistoxin, many of the most potent marine natural products will never be used as medicines, but may be of great importance as biochemical tools[13]. One example would be the palytoxin originally isolated in Hawaii from the tropical soft coral

*Corresponding author: Marisa Rangel, Immunopathology Laboratory, Division of Scientific Development, Butantan Institute, 05503-900, Sao Paulo, Brazil; Laboratory of Toxinology, Department of Physiological Sciences, University of Brasilia, 70910-900, Brasilia, DF, Brazil.

Tel/Fax: +551126279778

E-mail: marisarangel2112@gmail.com, marisa.rangel@butantan.gov.br

Foundation Project: Supported by CNPq (National Council for Scientific and Technological Development) (Grant No. 473645/2012-2), and received a postdoctoral fellowship from CAPES (Coordination for the Improvement of Higher Level - or Education- Personnel).

Palythoa sp., a zoanthid[14-16]. This substance and its analogs found in dinoflagellates from the genus *Ostreopsis* are very toxic to all mammal cells, due to the blocking of Na⁺/K⁺-ATPase pump, but are powerful tools to study cellular ion transport mechanisms[13,17].

Tetrodotoxin (TTX) and saxitoxin are guanidine neurotoxins found in many marine organisms, such as mussels and fishes, which bind to the site 1 of voltage dependent sodium channels, blocking the propagation of action potentials on muscle and nerve cells[18,19]. Okadaic acid, a lipophilic polyether originally isolated from the sponges *Halichondria okadai*, *Halichondria melanodocia* and from several dinoflagellates species[20,21], inhibits specifically cell phosphatases that regulate different cellular processes such as ion balance, neurotransmission and the cell cycle, being a well known tumor promoter[22,23].

These three toxins (TTX, saxitoxin and okadaic acid) are responsible for several cases of human poisoning, caused by the consumption of fish, clams, oysters and mussels that accumulated these toxins[24-26]. Despite that, due to its specific and reversible activity on voltage dependent sodium channels (Na_v 1.1-1.4, 1.6-1.7), two formulations containing TTX are currently under clinical trials.

In vitro cytotoxicity was the most prominent bioactivity searched in novel marine compounds for the past 30 years[27,28]. Therefore, besides cytarabine (Ara-C), many other compounds targeting cancer therapy are currently under investigation.

The Developmental Therapeutics Program of the U.S. National Cancer Institute (NCI) has a library of approximately 200 000 extracts from plants, marine organisms, bacteria and fungi. More than 13 000 marine animal specimens were processed, comprising 6 100 and 450 different species of marine animals and plants, respectively. From those, over 5 000 specimens belonged to the Phylum Porifera[29]. Results obtained from the NCI *in vitro* screening program (NCI60, the human tumor cell line assay) indicated that in late 1990's marine invertebrates were the major source of extracts with significant activity, with a probability to find an active extract near four times higher than in terrestrial animals and plants, microorganisms and marine algae. Amongst marine invertebrates, the sponges presented the strongest activity spectrum, followed by tunicates, cnidarians and bryozoans[30].

Since 1985, most novel marine bioactive compounds were isolated from invertebrates, Porifera and Cnidaria accounted for 56.89% of the total bioactive compounds[28]. Considering all articles on marine natural products published in the last 50 years, the most studied Phylum was the Porifera, although the proportion of new compounds described for sponges has diminished since the mid 1990s. Starting from this period, Ascomycota, Actinobacteria, Cyanobacteria and Cnidaria had an increase in the number of studies[8,31-33], while Rhodophyta, Ochrophyta, Echinodermata and Mollusca had a decrease in their popularity[34].

According to Hu *et al.*[28], only approximately 25% of more than 16 000 new marine natural products described since 1985 had their biological activities investigated. Additionally, the average proportion of bioactive compounds among the novel compounds is declining according to the data published in the last decade. These authors suggest that the technologies to find new compounds are more advanced than the research tools and methods for prospecting bioactivity[28]. Nonetheless, considering that more than 1 000 new molecules have been described from marine organisms yearly for the past eight years[34-41], the biotechnological and pharmaceutical

potential of the sea is definitively very impressive.

In the present review we focused on the marine natural products that are under clinical trials, or that have been already approved as drugs, grouped according to the therapeutic targets. Their chemical structures, mechanisms of action and a brief history of their discovery and development are presented.

2. Antitumor

Until now, four drugs based on marine natural products entered the market for cancer treatment, while nine are promising molecules in clinical trials. Their structures are represented in the Figures 1 and 2. The investigations of their mechanisms of action have shown them to be unique, in some cases. Promising antitumor drugs targeting apoptosis and the transcription factor NF-κB among other mechanisms have been reviewed[42,43].

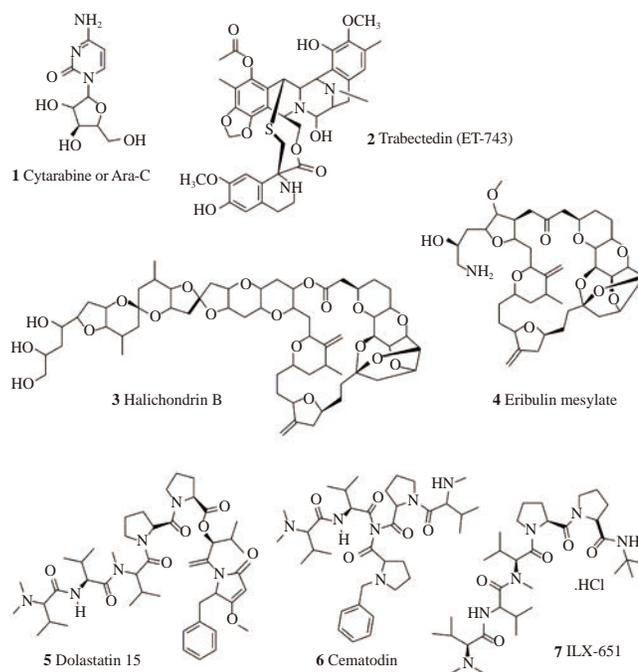


Figure 1. Marine-derived antitumoral drugs that are on the market (1-3) or in clinical trials. The sources of the natural compounds include sponges, tunicates and sea hares.

As mentioned above, the nucleosides from the Caribbean sponge *Tectitethya crypta*[1] were used as a model for the synthesis of analogs such as cytarabine or Ara-C **1**, the active substance of Cytosar-U® (Upjohn)[44]. Ara-C was approved for medicinal use in 1972, indicated for treatment of leukemia and lymphoma[4].

The marine tetrahydroisoquinoline alkaloid ecteinascidin 743 (generic name trabectedin, but also known as ET-743) **2**, produced by the tunicate *Ecteinascidia turbinata*, is a broad spectrum antitumor agent[2] first described by researchers from Illinois University (USA)[45], and licensed to PharmaMar (Spain) for pharmaceutical development. ET-743 interferes with cell division, and DNA transcription and repair mechanisms by binding in the minor groove of DNA molecules. The compound was obtained by synthesis in order to allow the sustainability of the production. From the isolation of the natural compound to the approval by Food and Drug Administration (FDA) and the European Agency for the Evaluation of Medicinal Products (EMA), it took almost 40 years[42]: Yondelis® is the trade name for the medicine developed with the semi-synthetic

drug and it was approved in 2007 for advanced or metastatic soft tissue sarcoma treatment, and in 2009 for therapy of ovarian cancer in association with pegylated liposomal doxorubicin DOXIL®/Caelyx® (www.pharmamar.com/yondelis.aspx). A review about trabectedin was published recently[46].

The macrocyclic polyether halichondrin B **3**[47] obtained from several species of marine sponges, such as *Halichondria okadai*, *Axinella* spp. and *Phakellia* spp., was also a model for the development of anticancer drugs[48,49]. The chemical synthesis of halichondrin B[50], enabled the discovery of the simplified analog, eribulin mesylate (E7389) **4**[51-53], approved in 2010 and commercialized as Halaven® for metastatic breast cancer chemotherapy[54]. Developed and marketed by Eisai Co (Japan), this compound affects microtubule polymerization, and therefore the mitosis (http://www.eisai.com/ir/individual/word/word_h03.html).

Dolastatins are potent antimetabolic polypeptides originally isolated from the sea hare *Dolabella auricularia* (Phylum Mollusca), and also produced by Cyanobacteria[55]. Different synthetic analogs of dolastatin-15 **5** were produced[56]. Cematodin (LU-103793) **6** was developed by ABBOTT GmbH & Co. KG (Germany)[57], and concluded phase II of clinical trials for melanoma. BASF Pharma synthesized tasidotin hydrochloride (ILX-651) **7** that was licensed for development to ILEX Oncology (USA) and concluded clinical trials (phase I) against advanced solid tumors (colorectal, lung, kidneys and pancreas)[58]. Dolastatin 10 **8** reached phase II of clinical trials[59], but was withdrawn. Years later, a dolastatin 10 synthetic analog,

monomethyl auristatin E **9**[60] was conjugated with an antibody and showed to be effective on cancer therapy, especially carcinoma[61]. The chimerical antibody conjugated with monomethyl auristatin E got the generic name “brentuximab vedotin” and was launched on market in 2011 by Seattle Genetics Inc. as ADCETRIS®, indicated for the treatment of lymphomas. Glembatumumab vedotin (CDX011), PSMA-ADC and ABT-414 are other antibody drug conjugates linked to the toxins monomethyl auristatin E and monomethyl auristatin F **10** that are currently in phase II of clinical trials for cancer treatment[62], under development by Celldex Therapeutics (<http://www.celldex.com/pipeline/cdx-011.php>), Progenics Pharmaceuticals (<http://www.progenics.com/product-pipeline/psma-adc-therapeutic-technology/>) and Abbvie (<http://www.abbvie.com/oncology/home-pipeline.html#>), respectively.

Didemnins are a family of cyclic depsipeptides isolated from the Caribbean tunicate *Trididemnum solidum*[63]. Besides antitumor activity, they presented potent antiviral properties in assays performed *in vitro* and *in vivo*. However, despite its important antiviral activity, didemnin B **11** has low selectivity and therapeutic index: it is cytotoxic and inhibits the synthesis of proteins, DNA and RNA of cells at the same concentrations that inhibit the virus growth. Starting in 1986, didemnin B underwent clinical trials (phase I and II)[30,48], but the tests were interrupted due to substantial toxic side effects[2].

In the decade of 1990, the depsipeptide dehydrididemnin B **12**, also known as aplidine or plitidepsin, was isolated from the

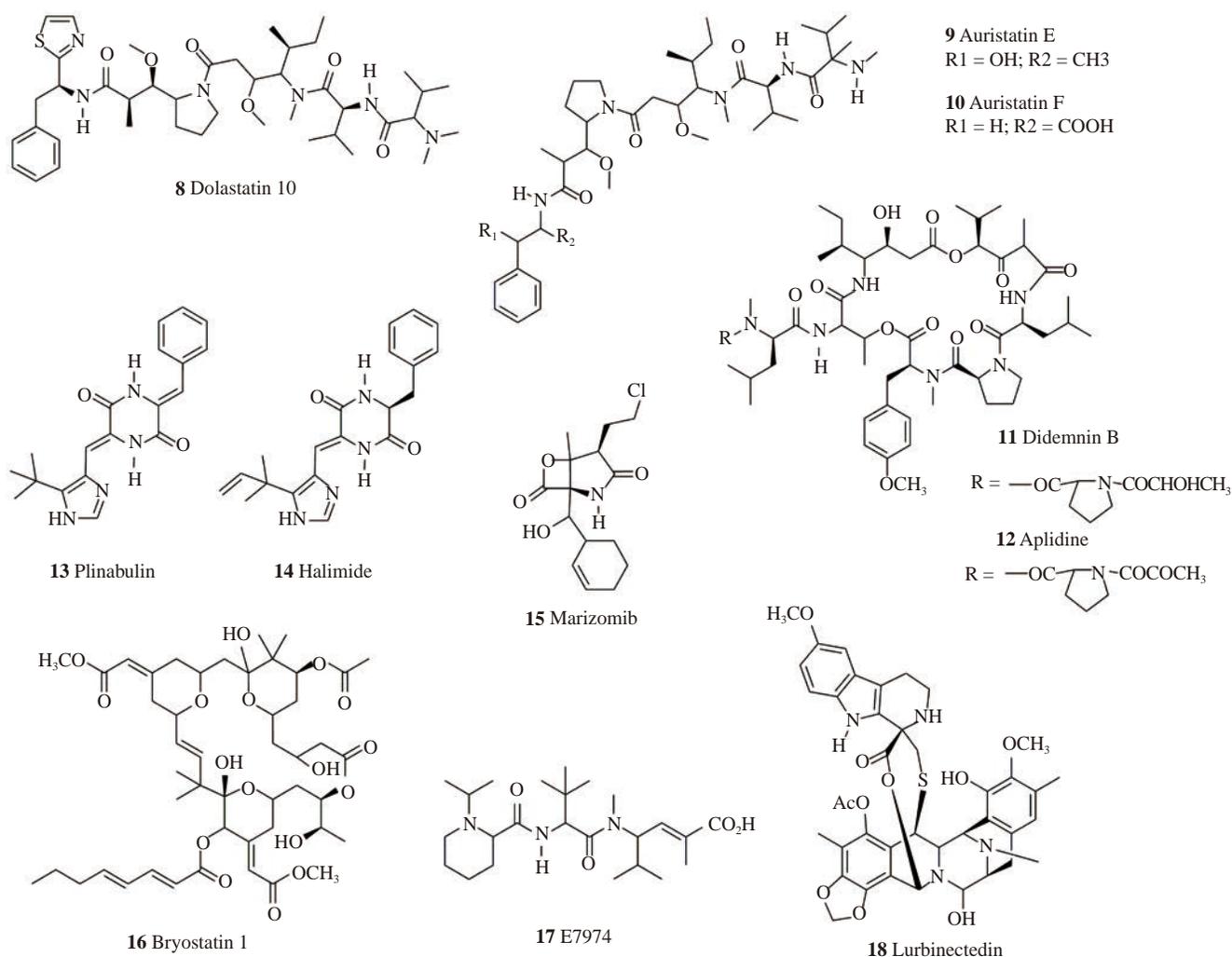


Figure 2. Structures of polypeptides, depsipeptides, diketopiperazines and other marine derived antitumoral drugs that are on the market (9) or clinical trials.

Mediterranean tunicate *Aplidium albicans*[64]. This substance has antiproliferative activity by blocking the cell cycle and inducing apoptosis, with strong activity against multiple myeloma cells[65-67]. PharmaMar is currently developing Aplidin® for the treatment of multiple myeloma (phase III of clinical trials), and for solid and hematological malignant neoplasias, like T-cell lymphoma (phase II of clinical trials) (<http://www.pharmamar.com/aplidin.aspx>).

Plinabulin (NPI-2358) **13**[8,68] is a synthetic analog of the diketopiperazine halimide (or phenylahistin) **14**[69], a natural product isolated from a marine fungi (*Aspergillus* sp.). Plinabulin inhibits tubulin polymerization, leading to the disruption of the vascular endothelial architecture of the tumor. This substance inhibits tumor growth through two different mechanisms: by inducing the collapse of the existing vasculature of the tumor, and promoting apoptosis. Phases I and II of clinical trials against solid tumors and lymphomas were conducted by the extinct Nereus Pharmaceuticals. Now, BeyondSpring Pharmaceuticals is developing plinabulin, and announced the start of phase III clinical trials in patients with non-small cell lung cancer for the first quarter of 2015 (<http://www.beyondspringpharma.com/press-release-plinabulin-phase-3-trial/>).

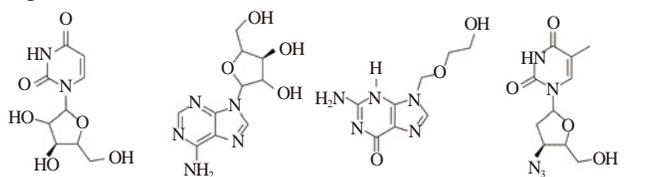
Triphase Accelerator Corporation is currently developing a medicine that contains marizomib (NPI-0052 or salinosporamide A) **15**, which is a natural product from the marine actinomycete *Salinispora tropicalis*[70], and a novel, highly potent proteasome inhibitor that irreversibly targets and inhibits all three proteasome subunits; it is active against cells resistant to proteasome inhibitor bortezomib[42]. Marizomib is on phase I of clinical trials against multiple myeloma, lymphomas, leukemias and solid tumors (<http://triphaseco.com/pipeline/>).

Other substances that have ongoing phase I clinical trials are: bryostatin **16**, from the bryozoan *Bugula neritina*[71]; E7974 **17**, an analog of the tripeptide hemiasterlin isolated from the sponges *Auleta* sp. and *Siphonochalina* sp., that inhibits microtubules polymerization[72,73]; and lurbinctedin or PM01183 **18**, an alkaloid related to the eictinascidins 2 that binds to certain DNA sequences, inducing apoptosis[74] (<http://www.pharmamar.com/pm01183-en.aspx>).

3. Antiviral

Marine natural products also played a very important role in the development of antiviral drugs. In a general way, there are more anticancer drugs (and a higher diversity of mechanisms of action) in the market than antiviral drugs. Nevertheless, the number of approved drugs of marine origin is similar in both classes.

Starting from the chemical structure of the nucleoside spongouridine **19**[1], semi synthetic modifications originated the antiviral drugs vidarabine or Ara-A/Vira A **20**, aciclovir **21**, and zidovudine [azidothymidine or AZT] **22**[2,75], that are shown in Figure 3.



19 Spongouridine **20** Vidarabine or Ara-A **21** Aciclovir **22** Zidovudine or AZT

Figure 3. The marine nucleoside spongouridine and their synthetic analogues used therapeutically as antiviral drugs.

Vidarabine, after being obtained by synthesis[76], has been produced by *Streptomyces antibioticus* fermentation to be marketed as Vira-A® since 1976. Vidarabine inhibits the DNA polymerases of herpes, vaccinia and varicella-zoster viruses[77]. It is also prescribed for the treatment of herpes virus related conjunctivitis[5]. Aciclovir (Zovirax®) is a guanosine analog indicated to treat infections caused by herpes simplex, zoster and varicella viruses[78], and is currently the first line drug for herpes simplex infections. In the market there are also some prodrugs of aciclovir, as valaciclovir[79].

AZT was first synthesized in 1964 by Jerome P. Horwitz as a potential anticancer compound. More than twenty years later the drug was found to be a potent antiretroviral drug by inhibiting the reverse transcriptase of HIV (RT-HIV)[80]. AZT was the first treatment available for HIV infected patients, prescribed under the trade name Retrovir®, and launched in 1987. The time lapse for the development of Retrovir® by Burroughs Wellcome (now GlaxoSmithKline), from the confirmation of its *in vitro* efficacy against HIV to its approval by the United States FDA, was only a little longer than two years (<http://www.fda.gov/ForPatients/Illness/HIVAIDS/History/ucm151074.htm>). Based on its efficacy and safety, several other nucleoside analogues were developed as RT-HIV inhibitors, like lamivudine (3TC), abacavir (ABC), etc. These substances still have a role as components of the antiretroviral therapy[81].

4. Analgesic

Some analgesic formulations derived from marine toxins were developed for the treatment of neuropathic and chronic pain, especially in morphine-unresponsive patients. One natural compound is already on the market (ziconotide) and another is on clinical trials (tetrodotoxin). Their structures are represented in Figure 4.

Neurex Corporation (Menlo Park, California, EUA) and Cognetix Inc. (Salt Lake City, Utah, EUA) synthesized the peptide ziconotide (ω -conotoxin MVIIA) **23**, obtained from the venom of the mussel *Conus magnus*. It is an N-type calcium channel blocker that reduces chronic and neuropathic pain[82,83]. This peptide (and others from *Conus* venom) was characterized by professors Baldomero Olivera (Utah University) and George Miljanich (California University), and it promotes the decrease of upper and lower limbs reflexes, thus reducing the spasticity caused by spinal cord injury[84]. Prialt® (Ziconotide) was launched in 2004 by Elan Pharmaceuticals as a new therapy for chronic pain due to its significant antinociceptive action even in morphine-unresponsive patients. It must be administered by intrathecal route, and therefore its use is restricted to hospitals[85-87]. Figure 4 presents the sequence of aminoacids and the disulfide bridges in its structure.

The guanidine alkaloid tetrodotoxin (TTX) **24**, a blocker of voltage dependent sodium channels isolated from fish, algae and bacteria[88], has shown therapeutic efficacy as analgesic in cancer patients. Two formulations are currently under evaluation in phases II and III of clinical trials by the Canadian WEX Pharmaceuticals Inc.: the first formulation is on phase III, indicated for the treatment of neuropathic pain in cancer patients, by intramuscular and subcutaneous administration; the second one is on phase II of clinical trials, for peripheral pain and cancer-related pain (http://www.wextech.ca/clinical_trials.asp?m=1&s=0&p=0; <http://www.clinicaltrials.gov>). A review on tetrodotoxin chemistry and toxicity

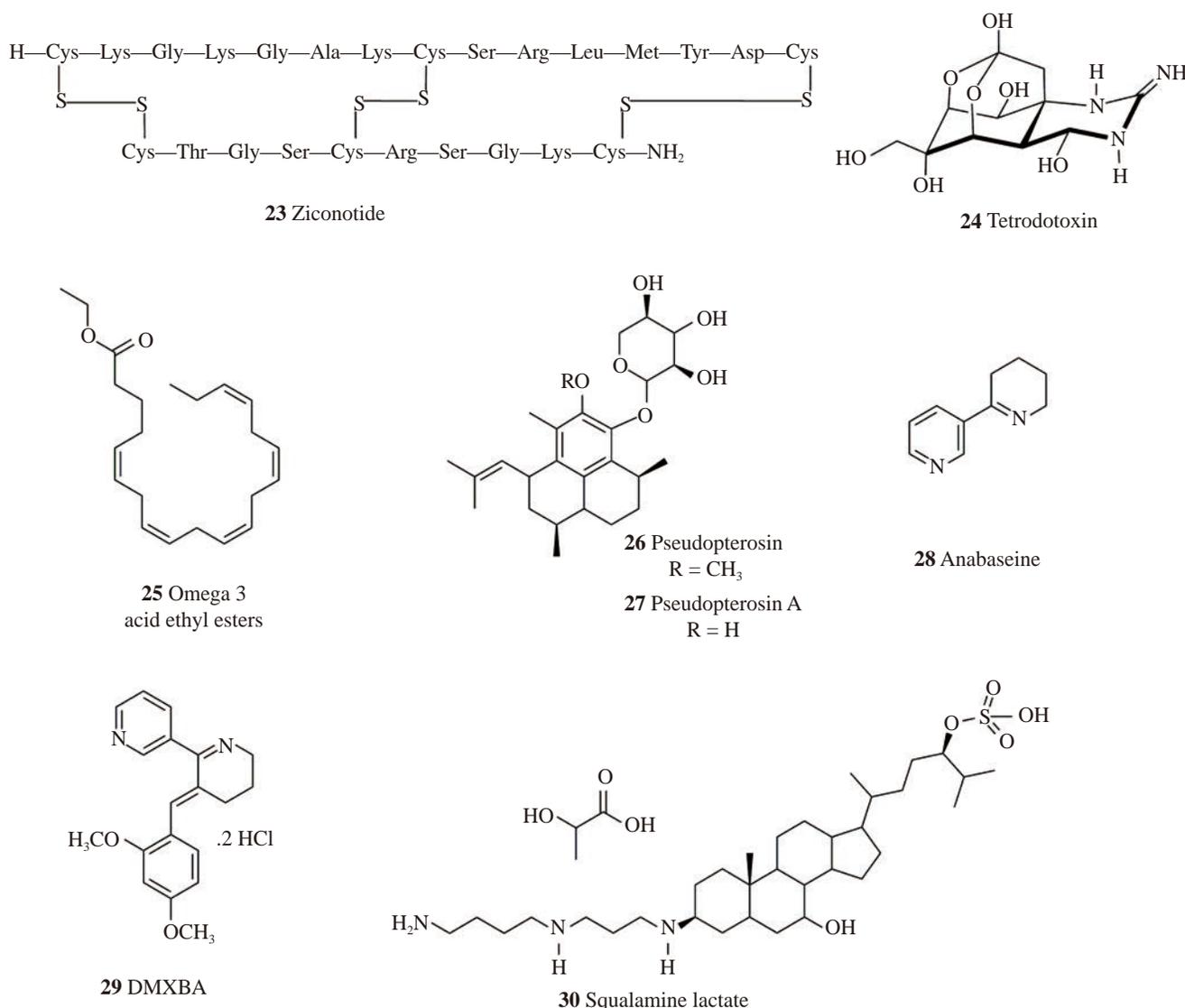


Figure 4. Marine natural products that are being commercialized as analgesic, for treatment of hypertriglyceridemia and marine derived drugs that are on clinical trials for other therapeutic uses.

was published recently[89].

5. Other applications

One marine derived drug was approved by the FDA to treat a metabolic disease, while three others are under clinical trials for miscellaneous applications. Their structures are represented on Figure 4.

In 2004 the FDA approved fish omega 3 acid ethyl esters (Lovaza®) **25** to treat hypertriglyceridemia[90,91], a very important risk factor for coronary heart disease. Lovaza® inhibits the triglyceride synthesizing enzymes, thus resulting in decreased synthesis and serum levels of triacylglycerols. These omega 3 esters, also commercialized outside USA as Omacor®, were developed by Reliant Pharmaceuticals, which was acquired by GlaxoSmithKline in 2007[92].

Pseudopterosin A methyl ether **26** (also known as VM301), a hemi synthetic derivative of pseudopterosin A **27**, a diterpene glycoside isolated from the soft coral *Pseudopterogorgia elizabethae*[93], presented anti-inflammatory and cicatrizing properties[49]. In double blind phase II clinical trials pseudopterosin increased reepithelization, and accelerated the wound healing process[5]. There are several reports of total synthesis of pseudopterosin analogues[94-

96], so one might expect new derivatives of pseudopterosin to be developed in the next years.

Anabaseine **28**[97] is a toxic alkaloid from several nemertean species, such as *Paranemertes peregrine* and *Amphiporus lactifloreus*[98], that seems to play ecological roles paralyzing preys, and as a feed-deterrent. Its synthetic derivative, DMXBA (3-(2,4-dimethoxybenzylidene)-anabaseine, also known as GTS-21 **29**, has the same mechanism as the natural compound: it stimulates the $\alpha 7$ nicotinic acetylcholine receptors expressed in neurons and astrocytes in central nervous system, and in the peripheral macrophages[99,100]. DMXBA improved cognition and sensory deficit in several animal models, and has shown neuroprotective effect *in vitro* and *in vivo*. Furthermore, presented anti-inflammatory activity mediated by $\alpha 7$ receptor in animal models. DMXBA (GTS-21) was licensed to Comentis Inc., which is developing drugs for treatment for Alzheimer's disease and schizophrenia on their clinical pipeline (<http://comentis.com/>). Phases I and II of clinical trials studies showed a significant cognitive improvement in healthy young adults and in schizophrenic patients[5]. Additional clinical trials (phases I and II) for schizophrenia therapy and other psychotic disorders are now under recruitment (<https://clinicaltrials.gov/ct2/results?term=DMXBA&Search=Search>).

Squalamine lactate (MSI-1256F) **30** is an amino sterol with antibiotic activity isolated from the stomach of the shark *Squalus acanthias*[101]. Squalamine is also a potent angiogenesis inhibitor, and thus was evaluated in several human clinical trials for cancer (<https://clinicaltrials.gov/ct2/results?term=Squalamine+lactate+&Search=Search>). OHR Pharmaceutical Inc. is currently developing MSI-1256F for ophthalmologic wet age-related macular degeneration (wet AMD), with ongoing clinical trials of phases II and III (<http://www.ohrpharmaceutical.com/research/squalamine>). Its potential in other ophthalmic problems involving pathological angiogenesis in the eye is also being evaluated[102].

6. Concluding remarks

The examples above illustrate briefly the history of marine natural products that originated medicines, or are in advanced clinical development. To date, clinical development of marine drugs focused mainly in cancer treatment[42,103], with at least nine derivatives currently under advanced clinical trials, and four already marketed. Most of the drugs presented in this review were first discovered as toxins, exerting ecological roles to the organisms that produce or accumulate them. Characterization of the mechanisms of action and the molecular targets of these compounds pointed them towards a possible pharmaceutical application. In general, in order to go further in the pharmaceutical industry pipeline these molecules must have low molecular weight, and their chemical synthesis elucidation and/or fermentation production are essential to guarantee the supply in adequate amount, since the harvesting of natural sources is usually scarce. In the last 25 years there was an increase in the number of bioactive natural products characterized from marine organisms, raising the expectation that many more medicines will be derived from marine natural products in the near future.

Conflict of interest statement

We declare that we have no conflict of interest.

Acknowledgments

The authors wish to thank Tauana Wanke for assistance with the figures.

Marisa Rangel is supported by CNPq (National Council for Scientific and Technological Development) (Grant 473645/2012-2), and received a postdoctoral fellowship from CAPES (Coordination for the Improvement of Higher Level -or Education- Personnel).

References

- [1] Bergmann W, Feeney RJ. Contributions to the study of marine products. XXXII. The nucleosides of sponges. I. *J Org Chem* 1951; **16**(6): 981-7.
- [2] Donia M, Hamann MT. Marine natural products and their potential applications as anti-infective agents. *Lancet Infect Dis* 2003; **3**(6): 338-48.
- [3] Ireland CM, Copp BR, Foster MD, McDonald LA, Radisky DC, Swersey JC. Biomedical potential of marine natural products. In: Attaway DH, Zaborsky OR, editors. *Marine biotechnology: pharmaceutical and bioactive natural products. Vol 1*. New York: Plenum Press; 1993, p. 1-43.
- [4] McConnell O, Longley RE, Koehn FE. *The discovery of natural products with therapeutic potential*. Boston (MA): Butterworth-Heinemann; 1994.
- [5] Mayer AM, Glaser KB, Cuevas C, Jacobs RS, Kem W, Little RD, et al. The odyssey of marine pharmaceuticals: a current pipeline perspective. *Trends Pharmacol Sci* 2010; **31**(6): 255-65.
- [6] Snoeck R, Andrei G, De Clercq E. Current pharmacological approaches to the therapy of varicella zoster virus infections: a guide to treatment. *Drugs* 1999; **57**(2): 187-206.
- [7] Oliveira JS, Freitas JC. [Marine natural products: food envenoming characteristics and substances of pharmacological interest]. *Hig Alimentar* 2001; **15**(20): 22-33. Portuguese.
- [8] Saleem M. Bioactive natural products from marine-derived fungi-an update. In: Atta-ur-Rahaman FRS, editor. *Studies in natural products chemistry. Vol 45*. Amsterdam: Elsevier; 2015, p. 297-361.
- [9] Cheer CJ, Pickles FJ. The crystal and molecular structure of 1,3-bis (thiocarbamoyl)-2-NN-dimethylaminopropane hydrochloride (Cartap). *J Chem Soc Perkin II* 1980; **2**(12): 1805-8.
- [10] Freitas JC. Biomedical importance of marine natural products. *Ciência e Cultura* 1990; **42**(1): 20-4.
- [11] Boorugu HK, Chrispal A. Cartap hydrochloride poisoning: a clinical experience. *Indian J Crit Care Med* 2012; **16**(1): 58-9.
- [12] Kumar ASP, Amalnath D, Dutta TK. Cartap poisoning: a rare case report. *Indian J Crit Care Med* 2011; **15**(4): 233-5.
- [13] Van Soest RWM, Van Kempen TMG, Braekman JC, editors. Sponges in time and space: biology, chemistry, paleontology. Proceedings of the 4th International Porifera Congress; 1994 Apr 19-23; Amsterdam, Netherlands. Rotterdam: A.A. Balkema Publishers; 1994.
- [14] Moore RE, Scheuer PJ. Palytoxin: a new marine toxin from a coelenterate. *Science* 1971; **172**(3982): 495-8.
- [15] Moore RE, Bartolini G. Structure of palytoxin. *J Am Chem Soc* 1981; **103**(9): 2491-4.
- [16] Uemura D, Ueda K, Hirata Y. Further studies on palytoxin. II. structure of palytoxin. *Tetrahedron Lett* 1981; **22**(29): 2781-4.
- [17] Ramos V, Vasconcelos V. Palytoxin and analogs: biological and ecological effects. *Mar Drugs* 2010; **8**(7): 2021-37.
- [18] Kao CY. Tetrodotoxin, saxitoxin and their significance in the study of excitation phenomena. *Pharmacol Rev* 1966; **18**(2): 997-1049.
- [19] Narahashi T. Mechanism of action of tetrodotoxin and saxitoxin on excitable membranes. *Fed Proc* 1972; **31**(3): 1124-32.
- [20] Tachibana K, Scheuer PJ, Tsukitani Y, Kikuchi H, Van Engen D, Clardy J, et al. Okadaic acid, a cytotoxic polyether from two marine sponges of the genus *Halichondria*. *J Am Chem Soc* 1981; **103**(9): 2469-71.
- [21] Yasumoto T, Oshima Y, Sugawara W, Fukuyo Y, Oguri H, Igarashi T, et al. Identification of *Dinophysis fortii* as the causative organism of diarrhetic shellfish poisoning. *Bull Jpn Soc Sci Fish* 1980; **46**: 1405-11.
- [22] Yasumoto T, Oshima Y, Yamaguchi M. Occurrence of a new type of shellfish poisoning in the Tohoku District. *Bull Jpn Soc Sci Fish* 1978; **44**: 1249-55.
- [23] Van Dolah FM. Marine algal toxins: origins, health effects, and their increased occurrence. *Environ Health Perspect* 2000; **108** (Suppl 1): 133-41.
- [24] Kacem I, Bouaïcha N, Hajjem B. Comparison of okadaic acid profiles in mussels and oysters collected in Mediterranean Lagoon, Tunisia. *Int J Biol* 2010; **2**(2): 238-45.
- [25] Faber S. Saxitoxin and the induction of paralytic shellfish poisoning. *J Young Invest* 2012; **23**(1): 1-7.
- [26] Cole JB, Heegaard WG, Deeds JR, McGrath SC, Handy SM. Tetrodotoxin poisoning outbreak from imported dried puffer fish-Minnesota, Minneapolis, 2014. *Morb Mortal Wkly Rep* 2015; **63**(51):

- 1222-5.
- [27] Blunt JW, Copp BR, Munro MH, Northcote PT, Prinsep MR. Marine natural products. *Nat Prod Rep* 2006; **23**(1): 26-78.
- [28] Hu Y, Chen J, Hu G, Yu J, Zhu X, Lin Y, et al. Statistical research on the bioactivity of new marine natural products discovered during the 28 years from 1985 to 2012. *Mar Drugs* 2015; **13**(1): 202-21.
- [29] McCloud TG. High throughput extraction of plant, marine and fungal specimens for preservation of biologically active molecules. *Molecules* 2010; **15**(7): 4526-63.
- [30] Munro MH, Blunt JW, Dumdei EJ, Hickfort SJ, Lill RE, Li S, et al. The discovery and development of marine compounds with pharmaceutical potential. *J Biotechnol* 1999; **70**(1-3): 15-25.
- [31] Tan LT. Bioactive natural products from marine cyanobacteria for drug discovery. *Phytochemistry* 2007; **68**(7): 954-79.
- [32] Manivasagan P, Kang KH, Sivakumar K, Li-Chan EC, Oh HM, Kim SK. Marine actinobacteria: an important source of bioactive natural products. *Environ Toxicol Pharmacol* 2014; **38**(1): 172-88.
- [33] Liu QA, Zheng JJ, Gu YC, Wang CY, Shao CL. The chemistry and bioactivity of macrolides from marine microorganisms. In: Atta-ur-Rahaman FRS, editor. *Studies in natural products chemistry. Vol 44*. Amsterdam: Elsevier; 2015, p. 353-401.
- [34] Blunt JW, Copp BR, Keyzers RA, Munro MH, Prinsep MR. Marine natural products. *Nat Prod Rep* 2015; **32**(2): 116-211.
- [35] Blunt JW, Copp BR, Hu WP, Munro MH, Northcote PT, Prinsep MR. Marine natural products. *Nat Prod Rep* 2008; **25**(1): 35-94.
- [36] Blunt JW, Copp BR, Hu WP, Munro MH, Northcote PT, Prinsep MR. Marine natural products. *Nat Prod Rep* 2009; **26**(2): 170-244.
- [37] Blunt JW, Copp BR, Munro MH, Northcote PT, Prinsep MR. Marine natural products. *Nat Prod Rep* 2010; **27**(2): 165-237.
- [38] Blunt JW, Copp BR, Munro MH, Northcote PT, Prinsep MR. Marine natural products. *Nat Prod Rep* 2011; **28**(2): 196-268.
- [39] Blunt JW, Copp BR, Keyzers RA, Munro MH, Prinsep MR. Marine natural products. *Nat Prod Rep* 2012; **29**(2): 144-222.
- [40] Blunt JW, Copp BR, Keyzers RA, Munro MH, Prinsep MR. Marine natural products. *Nat Prod Rep* 2013; **30**(2): 237-323.
- [41] Blunt JW, Copp BR, Keyzers RA, Munro MH, Prinsep MR. Marine natural products. *Nat Prod Rep* 2014; **31**(2): 160-258.
- [42] von Schwarzenberg K, Vollmar AM. Targeting apoptosis pathways by natural compounds in cancer: marine compounds as lead structures and chemical tools for cancer therapy. *Cancer Lett* 2013; **332**(2): 295-303.
- [43] Folmer F, Jaspars M, Dicato M, Diederich M. Marine natural products as targeted modulators of the transcription factor NF-kappaB. *Biochem Pharmacol* 2008; **75**(3): 603-17.
- [44] Talley RW, O'Bryan RM, Tucker WG, Loo RV. Clinical pharmacology and human antitumor activity of cytosine arabinoside. *Cancer* 1967; **20**(5): 809-16.
- [45] Rinehart KL, Holt TG, Fregeau NL, Keifer PA, Wilson GR, Perun TJ Jr, et al. Bioactive compounds from aquatic and terrestrial sources. *J Nat Prod* 1990; **53**(4): 771-92.
- [46] Galmarini CM, D'Incalci M, Allavena P. Trabectedin and plitidepsin: drugs from the sea that strike the tumor microenvironment. *Mar Drugs* 2014; **12**(2): 719-33.
- [47] Hirata Y, Uemura D. Halichondrins-antitumor polyether macrolides from a marine sponge. *Pure Appl Chem* 1986; **58**(5): 701-10.
- [48] Altmann KH. Microtubule-stabilizing agents: a growing class of important anticancer drugs. *Curr Opin Chem Biol* 2001; **5**(4): 424-31.
- [49] Proksch P, Edrada RA, Ebel R. Drugs from the seas-current status and microbiological implications. *Appl Microbiol Biotechnol* 2002; **59**(2-3): 125-34.
- [50] Aicher TD, Buszek KR, Fang FG, Forsyth CJ, Jung SH, Kishi Y, et al. Total synthesis of halichondrin B and norhalichondrin B. *J Am Chem Soc* 1992; **114**(8): 3162-4.
- [51] Kishi Y, Fang F, Forsyth CJ, Scola PM, Yoon SK, inventors; Harvard College, assignee. Halichondrins and related compounds. WO1993017690 A1. 1993 Sep 16.
- [52] Littlefield BA, Palme M, Seletsky BM, Towle MJ, Yu MJ, Zheng W, inventors; Eisai Co Ltd., Littlefield BA, Palme M, Seletsky BM, Towle MJ, Yu MJ, Zheng W, assignees. Macrocylic analogs and methods of their use and preparation. WO 9965894 A1. 1999 Dec 23.
- [53] Choi H, Demeke D, Kang FA, Kishi Y, Nakajima K, Nowak P, et al. Synthetic studies on the marine natural product halichondrins. *Pure Appl Chem* 2003; **75**(1): 1-17.
- [54] Liu KK, Sakya SM, O'Donnell CJ, Flick AC, Ding HX. Synthetic approaches to the 2010 new drugs. *Bioorg Med Chem* 2012; **20**(3): 1155-74.
- [55] Luesch H, Moore RE, Paul VJ, Mooberry SL, Corbett TH. Isolation of dolastatin 10 from the marine cyanobacterium *Symploca* species VP642 and total stereochemistry and biological evaluation of its analogue symplostatin 1. *J Nat Prod* 2001; **64**(7): 907-10.
- [56] Bai R, Friedman SJ, Pettit GR, Hamel E. Dolastatin 15, a potent antimetabolic depsipeptide derived from *Dolabella auricularia*. Interaction with tubulin and effects of cellular microtubules. *Biochem Pharmacol* 1992; **43**(12): 2637-45.
- [57] de Arruda M, Cocchiario CA, Nelson CM, Grinnell CM, Janssen B, Haupt A, et al. LU103793 (NSC D-669356): a synthetic peptide that interacts with microtubules and inhibits mitosis. *Cancer Res* 1995; **55**(14): 3085-92.
- [58] Ray A, Okouneva T, Manna T, Miller HP, Schmid S, Arthaud L, et al. Mechanism of action of the microtubule-targeted antimetabolic depsipeptide tasidotin (formerly ILX651) and its major metabolite tasidotin C-carboxylate. *Cancer Res* 2007; **67**(8): 3767-76.
- [59] Bai RL, Pettit GR, Hamel E. Binding of dolastatin 10 to tubulin at a distinct site for peptide antimetabolic agents near the exchangeable nucleotide and vinca alkaloid sites. *J Biol Chem* 1990; **265**(28): 17141-9.
- [60] Francisco JA, Cerveny CG, Meyer DL, Mixan BJ, Klussman K, Chace DF, et al. cAC10-vcMMAE, an anti-CD30-monomethyl auristatin E conjugate with potent and selective antitumor activity. *Blood* 2003; **102**(4): 1458-65.
- [61] Doronina SO, Toki BE, Torgov MY, Mendelsohn BA, Cerveny CG, Chace DF, et al. Development of potent monoclonal antibody auristatin conjugates for cancer therapy. *Nat Biotechnol* 2003; **21**(7): 778-84.
- [62] Doronina SO, Bovee TD, Meyer DW, Miyamoto JB, Anderson ME, Morris-Tilden CA, et al. Novel peptide linkers for highly potent antibody-auristatin conjugate. *Bioconjug Chem* 2008; **19**(10): 1960-3.
- [63] Rinehart KL Jr, Gloer JB, Cook JC Jr, Mizsak SA, Scahill TA. Structures of the didemmins, antiviral and cytotoxic depsipeptides from a Caribbean tunicate. *J Am Chem Soc* 1981; **103**(7): 1857-9.
- [64] Sakai R, Rinehart KL, Kishore V, Kundu B, Faircloth G, Gloer JB, et al. Structure-activity relationship of the didemmins. *J Med Chem* 1996; **39**(14): 2819-34.
- [65] Urdiales JL, Morata P, Núñez De Castro I, Sánchez-Jiménez F. Antiproliferative effect of dehydroididemnin B (DDB), a depsipeptide isolated from Mediterranean tunicates. *Cancer Lett* 1996; **102**(1-2): 31-7.
- [66] Depenbrock H, Peter R, Faircloth GT, Manzanares I, Jimeno J, Hanauke AR. *In vitro* activity of aplidine, a new marine-derived anticancer compound, on freshly explanted clonogenic human tumor cells

- and hematopoietic precursor cells. *Br J Cancer* 1998; **78**(6): 739-44.
- [67] Mitsiades CS, Ocio EM, Pandiella A, Maiso P, Gajate C, Garayoa M, et al. Aplidin, a marine organism-derived compound with potent antimyeloma activity *in vitro* and *in vivo*. *Cancer Res* 2008; **68**(13): 5216-25.
- [68] Nicholson B, Lloyd GK, Miller BR, Palladino MA, Kiso Y, Hayashi Y, et al. NPI-2358 is a tubulin-depolymerizing agent: *in-vitro* evidence for activity as a tumor vascular-disrupting agent. *Anticancer Drugs* 2006; **17**(1): 25-31.
- [69] Fenical W, Jensen PR, Cheng XC, inventors; The Regents of the University of California, assignee. Halimide, a cytotoxic marine natural product, and derivatives thereof. United States Patent 6069146. 2000 May 30.
- [70] Feling RH, Buchanan GO, Mincer TJ, Kauffman CA, Jensen PR, Fenical W. Salinosporamide A: a highly cytotoxic proteasome inhibitor from a novel microbial source, a marine bacterium of the new genus salinospora. *Angew Chem Int Ed Engl* 2003; **42**(3): 355-7.
- [71] Pettit GR, Herald CL, Doubek DL, Herald DL, Arnold E, Clardy J. Isolation and structure of Bryostatin 1. *J Am Chem Soc* 1982; **104**: 6846-8.
- [72] Kuznetsov G, TenDyke K, Towle MJ, Cheng H, Liu J, Marsh JP, et al. Tubulin-based antimetabolic mechanism of E7974, a novel analogue of the marine sponge natural product hemiasterlin. *Mol Cancer Ther* 2009; **8**: 2852-60.
- [73] Rocha-Lima CM, Bayraktar S, Macintyre J, Raez L, Flores AM, Ferrell A, et al. A phase 1 trial of E7974 administered on day 1 of a 21-day cycle in patients with advanced solid tumors. *Cancer* 2012; **118**(17), 4262-70.
- [74] Leal JFM, Martínez-Díez M, García-Hernández V, Moneo V, Domingo A, Bueren-Calabuig JA, et al. PM01183, a new DNA minor groove covalent binder with potent *in vitro* and *in vivo* anti-tumour activity. *Br J Pharmacol* 2010; **161**: 1099-110.
- [75] Tziveleka LA, Vagias C, Roussis V. Natural products with anti-HIV activity from marine organisms. *Curr Top Med Chem* 2003; **3**: 1512-35.
- [76] Lee WW, Benitez A, Goodman L, Baker BR. Potential anticancer agents. I. synthesis of the β -anomer of 9-(d-arabinofuranosyl)-adenine. *J Am Chem Soc* 1960; **82**(10): 2648-9.
- [77] Whitley RJ, Tucker BC, Kinkel AW, Barton NH, Pass RF, Whelchel JD, et al. Pharmacology, tolerance, and antiviral activity of vidarabine monophosphate in humans. *Antimicrob Agents Chemother* 1980; **18**(5): 709-15.
- [78] Schaeffer HJ, Beauchamp L, de Miranda P, Elion GB, Bauer DJ, Collins P. 9-(2-Hydroxyethoxymethyl) guanine activity against viruses of the herpes group. *Nature* 1978; **272**: 583-5.
- [79] Zhang Y, Gao Y, Wen X, Ma H. Current prodrug strategies for improving oral absorption of nucleoside analogues. *Asian J Pharm Sci* 2014; **9**: 65-74.
- [80] Mitsuya H, Weinhold KJ, Furman PA, St Clair MH, Lehrman SN, Gallo RC, et al. 3'-Azido-3'-deoxythymidine (BW A509U): an antiviral agent that inhibits the infectivity and cytopathic effect of human T-lymphotropic virus type III/lymphadenopathy-associated virus *in vitro*. *Proc Natl Acad Sci U S A* 1985; **82**(20): 7096-100.
- [81] World Health Organization. Consolidated guidelines on the use of antiretroviral drugs for treating and preventing HIV infection: recommendations for a public health approach. Geneva: World Health Organization; 2013. [Online] Available from: <http://www.who.int/hiv/pub/guidelines/arv2013/download/en/> [Accessed on 15th March, 2015]
- [82] Olivera BM, Cruz LJ, de Santos V, LeCheminant GW, Griffin D, Zeikus R, et al. Neuronal calcium channel antagonists. Discrimination between calcium channel subtypes using omega-conotoxin from *Conus magus* venom. *Biochemistry* 1987; **26**(8): 2086-90.
- [83] Miljanich GP. Ziconotide: neuronal calcium channel blocker for treating severe chronic pain. *Curr Med Chem* 2004; **11**(23): 3029-40.
- [84] Olivera BM, Cruz LJ. Conotoxins, in retrospect. *Toxicon* 2001; **39**: 7-14.
- [85] Snutch TP. Targeting chronic and neuropathic pain: the N-type calcium channel comes of age. *NeuroRx* 2005; **2**: 662-70.
- [86] McGivern JG. Ziconotide: a review of its pharmacology and use in the treatment of pain. *Neuropsychiatr Dis Treat* 2007; **3**: 69-85.
- [87] Hannon HE, Atchison WD. Omega-conotoxins as experimental tools and therapeutics in pain management. *Mar Drugs* 2013; **11**: 680-99.
- [88] Yasumoto T, Yotsu M, Endo A, Murata M, Naoki H. Interspecies distribution and biogenetic origin of tetrodotoxin and its derivatives. *Pure Appl Chem* 1989; **61**: 505-8.
- [89] Bane V, Lehane M, Dikshit M, O'Riordan A, Furey A. Tetrodotoxin: chemistry, toxicity, source, distribution and detection. *Toxins* 2014; **6**: 693-755.
- [90] Bays H. Clinical overview of Omacor: a concentrated formulation of omega-3 polyunsaturated fatty acids. *Am J Cardiol* 2006; **98**: 71i-76i.
- [91] Davidson MH, Stein EA, Bays HE, Maki KC, Doyle RT, Shalwitz RA, et al. Efficacy and tolerability of adding prescription omega-3 fatty acids 4 g/d to Simvastatin 40 mg/d in hypertriglyceridemic patients: an 8-week, randomized, double-blind, placebo-controlled study. *Clin Ther* 2007; **29**(7): 1354-67.
- [92] Koski RR. Omega-3-acid ethyl esters (Lovaza) for severe hypertriglyceridemia. *Pharm Ther* 2008; **33**(5): 271-303.
- [93] Look SA, Fenical W, Jacobst RS, Clardy J. The pseudoopterosins: anti-inflammatory and analgesic natural products from the sea whip *Pseudoopterosorgia elisabethae*. *Proc Natl Acad Sci USA* 1986; **83**: 6238-40.
- [94] Coates RM, Kem WR, Abbott BC. Isolation and structure of a hoplonemertine toxin. *Toxicon* 1971; **9**: 15-22.
- [95] Buszek KR, Bixby DL. Total synthesis of pseudoopterosin A and E aglycon. *Tetrahedron Letters* 1995; **36**: 9129-32.
- [96] Flachsman F, Schellhaas K, Moya CE, Jacobs RS, Fenical W. Synthetic pseudoopterosin analogues: a novel class of antiinflammatory drug candidates. *Bioorg Med Chem* 2010; **18**: 8324-33.
- [97] McCulloch MWB, Kerr RG. Rapid structural diversification of pseudoopterosins: sulfuric acid promoted dehydro-aromatization yielding 14,15-dihydro-elisabatin B. *Tetrahedron Letters* 2015; **56**: 2030-33.
- [98] Kem WR. A study of the occurrence of anabaseine in *Paranemertes* and other nemertines. *Toxicon* 1971; **9**: 23-32.
- [99] Kem WR. The brain alpha7 nicotinic receptor may be an important therapeutic target for the treatment of Alzheimer's disease: studies with DMXBA (GTS-21). *Behav Brain Res* 2000; **113**: 169-81.
- [100] Kem WR, Soti F, Wildeboer K, LeFrancois S, MacDougall K, Wei DQ, et al. The nemertine toxin anabaseine and its derivative DMXBA (GTS-21): chemical and pharmacological properties. *Mar Drugs* 2006; **4**: 255-73.
- [101] Moore KS, Wehrli S, Rodert H, Rogers M, Forrest JN Jr, McCrimmon D, et al. Squalamine: an aminosterol antibiotic from the shark. *Proc Natl Acad Sci U S A* 1993; **90**: 1354-8.
- [102] Sene A, Chin-Yee D, Apte RS. Seeing through VEGF: innate and adaptive immunity in pathological angiogenesis in the eye. *Trends Mol Med* 2015; **21**: 43-51.
- [103] Kittakoop P. Anticancer drugs and potential anticancer leads inspired by natural products. In: Atta-ur-Rahaman, editor. *Studies in natural products chemistry*. Amsterdam: Elsevier; 2015, p. 251-307.