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Synchronization of ovulation in goats using prostaglandin F2 α based protocols during the breeding season

João Simões*

Department of Veterinary Science, University of Trás-os-Montes e Alto Douro, 5000-811 Vila Real, Portugal

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ABSTRACT

The main aim of this review was to describe the feasibility of prostaglandin F2 α (PGF2 α) based protocols used as a tool for ovulation synchronization in cycling goats. There is a reproductive seasonality in small ruminants. However, from latitudes 45° towards equator, the intensity of anoestrus progressively decreases and tends to disappear in local breeds. Consequently, PGF2 α or their synthetic analogues as luteolytic substances can assume a great importance in reproductive management of flocks from these regions. However, a single or double (9–11 days apart) PGF2 α administration provokes a good induction but a moderate synchronization of ovulations if timed artificial insemination is considered, and a significant short oestrous cycle can occur with detrimental effects on fertility rate when compared with conventional progesterone-based protocols. In order to minimize this constraint, some gonadotropin-releasing hormone-PGF2 α -gonadotropin-releasing hormone (OvSynch)-based protocols and their modifications, manipulating the dominant follicles and corpora lutea, were successfully tested in goats. Similar to cows, these PGF2 α based protocols seem to be a promising and more cost-effective tool for reproductive management in cycling goats.

1. Introduction

Goats and ewes show a marked reproductive seasonality mainly due to a photoperiod effect. The annual variation of the daily luminosity duration is perceived by the retina of these animals and a neurohormonal pathway involving suprachiasmatic nucleus, sympathetic neurons, and the pineal gland was responsible for production of melatonin, a key hormone, during short days/darkness periods[1-3]. Consequently, melatonin stimulates a neuronal network which modulates the hypothalamic secretion of gonadotropin-releasing hormone (GnRH)[3], and the hypothalamic-pituitary-gonadal axis is stimulated. From a latitude > 25°, most of the local breeds progressively present an anoestrous period with variations in duration (3 to 7–8 months) and intensity (originating up to 100% of

anoestrous females), during the natural long days[4,5].

In order to inseminate females out of season, several hormonal, natural or mixed protocols were developed for estrus and ovulation synchronization. These protocols were recently reviewed[6]. However, a demand of artificial inseminations also occurs during breeding season. Moreover, from regions at latitude less than 45° towards the equator, a progressive percentage of females remain (Spain; latitude 37° N) or can remain (Mexico; latitude 26° N) cyclic during whole year[5,7]. In fact, parturitions of goats can occur in all months under natural circumstances of mating (Figure 1).

Although classical hormonal protocol used during anoestrous season can also synchronize ovulations during the breeding season, other protocols more cost-effective can be used in cyclic goats during the breeding season, mainly in regions showing less marked seasonal reproductive activity of small ruminants.

The main aim of the present review was to describe useful methods for ovulation synchronization in goats, based on prostaglandin F2 α (PGF2 α) protocols during breeding season and to suggest the pathways for a more rational use of hormones for

*Corresponding author: João Simões, Department of Veterinary Science, University of Trás-os-Montes e Alto Douro, 5000-811 Vila Real, Portugal.

Tel: +351259350666

Fax: +351259350480

E-mail: jsimoes@utad.pt

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reproductive management in goats industry.

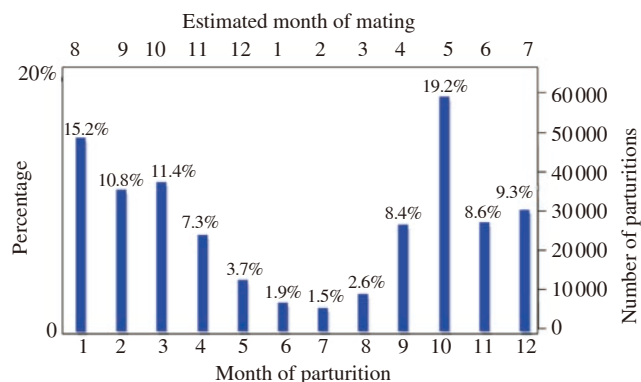


Figure 1. Monthly distribution of natural parturitions ($n = 292950$) after natural mating in Serrana goats (an indigenous Portuguese breed) from 1987 to 2015 (mainly at latitude 39° N– 41° N), according to the data obtained at Ruralbit (online Genpro® software; www.ruralbit.pt/). Data of abortion was removed. Estimation of natural mating (month) = month of parturition – 5 months (Unpublished data).

2. The classical hormonal protocol for ovulation synchronization in flocks

The classical hormonal protocol in goats, during non-breeding and breeding season, is based on a 11-day progesterone (P4) or their synthetic analogs (progestagens) priming + equine chorionic gonadotropin (eCG) + PGF2 α substances followed by a timed artificial insemination (TAI) with frozen or refrigerated straws 43 to 46 h after sponge withdrawal[4,8].

Progestagens modulate the pituitary luteinizing hormone secretion, inducing a negative feedback, modifying the hypothalamic GnRH activity[9], followed by a preovulatory luteinizing hormone surge after device withdrawal, in order to improve the development of follicles, including the preovulatory follicle(s), in females. In anoestrous goats, the eCG or a similar effect (such as male effect) is absolutely necessary for preovulatory follicles development. At time of progestagen/P4 device withdrawal or before 24–48 h[10], PGF2 α or

their synthetic analogues and eCG are administered intramuscularly in order to promote the quickly functional luteolysis of the corpora lutea (if present), and the development of antral (preovulatory) follicle(s), respectively. The occurrence of the luteolysis is crucial for ovulation occurrence in goats that present corpora lutea.

Leyva *et al.* observed that the progestagen/P4 priming increased the number of follicles stimulated by eCG and consequently the ovulation rate in anestrous ewes[11]. Moreover, the progestagen priming can synchronize the ovulatory wave[12]. At 5 to 7 days, short progestogen priming protocols were used, at least in some regions, in order to rationalize the use of progestagens or exogenous P4 without fertility decrease[12,13].

The exogenous P4 also can play an important role in order to improve fertility in cyclic females. This aspect was well evidenced in high lactating dairy cows. In fact, subluteal levels of endogenous P4, due to their high hepatic metabolism provoked by an increase of hepatic blood flux (greater ingestion of dry matter), appear to be a major cause of fertility decrease in this species[14].

Probably due to the expected fertility rate more than 50%, the usefulness of conventional protocols and the well-defined anoestrus season in countries located at high latitudes, progestagens/P4 based protocols were highly used in flock reproductive management. However, PGF2 α based protocols can be more used during breeding season, at least in regions at latitude $< 45^{\circ}$, followed by (timed) artificial insemination or natural mating.

3. Use of PGF2 α based protocols

Like cows, the induction of estrus/ovulation can be made in cycling ewes and goats with PGF2 α or their synthetic analogues by a single administration or two administrations 9–11 days apart[15,16]. In this last case, all irresponsive females at proestrus, estrus or early metaestrus (up to 2–3 days after ovulation) to first administration can be also synchronized at the second administration time.

The PGF2 α induced ovulation is effective under different

Table 1

Estrus, time of ovulation and fertility rate after single or double administration of PGF2 α in goats.

PGF2 α protocol	Estrus response	Onset of behavioural estrus ^a (h)	Time of ovulation ^a (h)	Fertility rate	References
Cloprostenol 100 μ g [single administration intramuscularly ($n = 30$)] on:	Early luteal phase ¹	100%	43.4 \pm 3.2	-	-
	Mid-luteal phase	80%	57.0 \pm 2.6	-	[18]
	Late luteal phase	80%	56.7 \pm 2.7	-	-
Cloprostenol 30 μ g (two doses, 10 days apart into vulvar submucosa)	79% (11/14)	47.7 \pm 10.1	87.5 \pm 12.8	55% (6/11) ^b	[19]
PGF2 α 7.5 mg (double intramuscular injection 12 days apart)	95% ($n = 38$)	59.5 \pm 4.2	-	80% (16/20) ^c	[20]
Cloprostenol 37.5 μ g (double intramuscular injection 10 days apart)	100% (14/14)	36.0 \pm 1.2	66.0 \pm 2.7	78% (11/14) ^c	[21]

^a: After (last) PGF2 α administration; ^b: Fertilization (frozen-thawed semen 16 to 18 h after the onset of estrus); ^c: Fertilization (natural mating); ¹: The early luteal phase showed a tendency ($P = 0.07$) for a higher estrus response percentage and for an early ($P < 0.01$) onset of behavioural estrus in goats than the mid- or late luteal phases.

environmental-nutritive conditions[17]. Normally, the estrus occurs 22 to 78 h after PGF2 α administration (Table 1), but the synchronization of the ovulation considering all goats is more disperse than P4 priming protocols. Bowdridge *et al.* suggested that two injections of prostaglandin separated by a period of 14 days can aid to synchronize estrus and ovulation[22]. However, the rate of response is high, reaching 90%–100%, for example, the PGF2 α alone promotes a good induction but a moderate synchronization of ovulation in flocks for TAI.

In order to minimize this problem, the detection of estrus should be implemented and a single TAI should be performed 12 h or even 16 to 18 h after the onset of estrus[23].

Gonzalez-Bulnes *et al.* suggested that the better response to PGF2 observed during the early than middle or later oestrus cycle phases is due to the lack of ovarian follicular waves synchronization in goats regarding the previous estrus phase, when normally a new follicular wave emerge[18]. However, no difference on periovulatory events were observed at latitude 3° N when cyclic goats were synchronized with the administration of 125 μ g of cloprostenol or the application of a controlled internal release device impregnated with 0.3 g of P4 suggesting that this PGF2 α based single protocol can be useful in their region[24].

In order to reduce the PGF2 α dose, Duque-Bonisolini *et al.* at latitude 10° N observed that the percentage of goats in estrus and the interval between treatments to onset of estrus remained similar when 87.50, 43.75 or 26.25 μ g of cloprostenol were administered intramuscularly[25]. These researchers also evidenced that the subsequent corpora lutea generated were similar among all three groups.

A major limitation of the PGF2 α use is the surge of short oestrous cycles and consequentially a detrimental effect on fertility. A lower fertility rate of PGF2 α than P4 sponges protocol (67% vs. 80%) was observed by Kusina *et al.*[26], and this can be due to deficiencies in the growth and functionality of luteal tissue[27]. In fact, a greater limitation of PGF2 α , used mainly without P4, priming is the occurrence of short estrus cycles (luteal insufficiency)[28]. It also shows in superovulated goats; this early luteal regression is caused by the premature release of endogenous PGF2 α and can be minimized by use of progestagens/P4[29,30].

Meanwhile, the GnRH–PGF2 α –GnRH (OvSynch)-based protocols given on Day 0, 7 and 9, respectively, or their subsequent modifications with and without P4 priming similar to those used on cows, were tested with success in goats[14,31,32]. The OvSynch protocol is based on the premise that the first GnRH administration can control the evolution of a healthy dominant follicle before the induced luteolysis of the corpora lutea[33]. The OvSynch protocol reached a pregnancy rate of 58% (14/24) when TAI was performed 16 h after the second GnRH administration and 66% (6/10) after natural mating in goats[21,31].

Bowdridge *et al.* reported that the NCSynch-TAI treatment protocol (PGF2 α –GnRH–PGF2 α –GnRH) is based on an OvSynch protocol plus a presynchronization treatment using PGF2 α with TAI (thawed frozen semen) at 72 h after the second PGF2 α administration (Figure 2)[22]. The proportion of goats ($n = 66$) in estrus was 73%. Moreover, all 66 goats were inseminated with frozen semen and the overall pregnancy rate was 68% (45/66). This protocol is similar to some Presynch-OvSynch protocols used in cows, which aimed to induce the presence of a mature dominant follicle at onset of the OvSynch treatment.

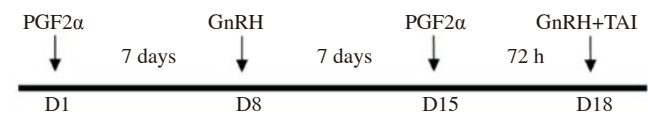


Figure 2. Schematic representation of the NCSynch-TAI protocol in goats[22].

These preliminary results indicate that the OvSynch protocols or their modifications are promising to synchronize ovulations in goats.

4. Conclusion

PGF2 α based protocols and their evolution, without P4 or progestagens sponge can be used in cycling goats during the breeding season, reaching reproductive indexes similar to the conventional protocols. In consequence, the reproductive management of flock can be more economical and improve the green production.

Conflict of interest statement

I declare that I have no conflict of interest.

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References

- [1] Ebling FJP, Hastings MH. The neural basis of seasonal reproduction. *Ann Zootech* 1992; **41**(3-4): 239-46.
- [2] Malpoux B, Thiéry JC, Chemineau P. Melatonin and the seasonal control of reproduction. *Reprod Nutr Dev* 1999; **39**(3): 355-66.
- [3] Malpoux B, Migaud M, Tricoire H, Chemineau P. Biology of mammalian photoperiodism and the critical role of the pineal gland and melatonin. *J Biol Rhythms* 2001; **16**(4): 336-47.
- [4] Leboeuf B, Delgadillo JA, Manfredi E, Piacère A, Clément V, Martin P, et al. Management of goat reproduction and insemination for genetic improvement in France. *Reprod Domest Anim* 2008; **43**(Suppl 2): 379-85.

- [5] Gallego-Calvo L, Gatica MC, Guzmán JL, Zarazaga LA. Role of body condition score and body weight in the control of seasonal reproduction in Blanca Andaluza goats. *Anim Reprod Sci* 2014; **151**(3-4): 157-63.
- [6] Simões J. Recent advances on synchronization of ovulation in goats, out of season, for a more sustainable production. *Asian Pac J Reprod* 2015; **4**(2): 157-65.
- [7] Delgadillo JA, Flores JA, Hernández H, Poindron P, Keller M, Fitz-Rodríguez G, et al. Sexually active males prevent the display of seasonal anestrus in female goats. *Horm Behav* 2015; **69**: 8-15.
- [8] Leboeuf B, Manfredi E, Boue P, Piacére A, Brice G, Baril G, et al. Artificial insemination of dairy goats in France. *Livest Prod Sci* 1998; **55**(3): 193-203.
- [9] Hansel W, Convey EM. Physiology of the estrous cycle. *J Anim Sci* 1983; **57**(Suppl 2): 404-24.
- [10] Baril G, Saumande J. Hormonal treatments to control time of ovulation and fertility of goats. In: Gruner L, Chabert Y, editors. Proceedings of the 7th International Conference on Goats; 2000 May 15-21; Tours, France. Tours: Institut de l'élevage; 2000, p. 400-5.
- [11] Leyva V, Buckrell BC, Walton JS. Regulation of follicular activity and ovulation in ewes by exogenous progestagen. *Theriogenology* 1998; **50**(3): 395-416.
- [12] Rubianes E, Menchaca A. The pattern and manipulation of ovarian follicular growth in goats. *Anim Reprod Sci* 2003; **78**(3-4): 271-87.
- [13] Menchaca A, Rubianes E. Pregnancy rate obtained with short-term protocol for timed artificial insemination in goats. *Reprod Domest Anim* 2007; **42**(6): 590-3.
- [14] Wiltbank MC, Souza AH, Carvalho PD, Bender RW, Nascimento AB. Improving fertility to timed artificial insemination by manipulation of circulating progesterone concentrations in lactating dairy cattle. *Reprod Fertil Dev* 2011; **24**(1): 238-43.
- [15] Greyling JPC. The effect of dose of prostaglandin in the synchronization of oestrus in the Boer goat doe. In: Proceedings of the Sixth International Conference in Goats; 1996; Beijing, China. Beijing: International Academic Publisher; 1996, p. 826.
- [16] Wiltbank MC, Pursley JR. The cow as an induced ovulator: timed AI after synchronization of ovulation. *Theriogenology* 2014; **81**(1): 170-85.
- [17] Khanum SA, Hussain M, Kausar R. Manipulation of estrous cycle in dwarf goat (*Capra hircus*) using estrumate under different management conditions. *Anim Reprod Sci* 2006; **92**(1-2): 97-106.
- [18] Gonzalez-Bulnes A, Díaz-Delfa C, Garcia-Garcia RM, Urrutia B, Carrizosa JA, Lopez-Sebastian A. Origin and fate of preovulatory follicles after induced luteolysis at different stages of the luteal phase of the oestrous cycle in goats. *Anim Reprod Sci* 2005; **86**(3-4): 237-45.
- [19] Esteves LV, Brandão FZ, Cruz RC, Souza JMG, Oba E, Facó O, et al. Reproductive parameters of dairy goats submitted to estrus synchronization with prostaglandin F2 associated or not to hCG at estrous onset. *Arq Bras Med Vet Zootec* 2013; **65**(6): 1585-92.
- [20] Amarantidis I, Karagiannidis A, Saratsis P, Brikas P. Efficiency of methods used for estrous synchronization in indigenous Greek goats. *Small Rumin Res* 2004; **52**(3): 247-52.
- [21] Riaz H, Sattar A, Arshad MA, Ahmad N. Effect of synchronization protocols and GnRH treatment on the reproductive performance in goats. *Small Rumin Res* 2012; **104**(1-3): 151-5.
- [22] Bowdridge EC, Knox WB, Whisnant CS, Farin CE. NCSynch: a novel, progestagen-free protocol for ovulation synchronization and timed artificial insemination in goats. *Small Rumin Res* 2013; **110**: 42-5.
- [23] Al Yacoub AN, Gaulty M, Sohnrey B, Holtz W. Fixed-time deep uterine insemination in PGF2 α -synchronized goats. *Theriogenology* 2011; **76**(9): 1730-5.
- [24] Bukar MM, Yusoff R, Ariff OM, Haron AW, Dhaliwal GK, Naing SW, et al. Comparison of the patterns of antral follicular development between hormonally synchronized and natural estrous cycles of non-seasonal, polyestrous goats in the tropics. *Reprod Biol* 2012; **12**(3): 325-8.
- [25] Duque-Bonisolí C, Salvador A, Díaz T, Contreras-Solis I. Ovarian response to oestrous synchronization protocol based on use of reduced doses of cloprostenol in cyclic goats. *Reprod Domest Anim* 2012; **47**(6): e79-82.
- [26] Kusina NT, Tarwirei, Hamudikuwanda H, Agumba G, Mukwena J. A comparison of the effects of progesterone sponges and ear implants, PGF2 α , and their combination on efficacy of estrus synchronization and fertility of Mashona goat does. *Theriogenology* 2000; **53**: 1567-80.
- [27] Vázquez MI, Blanch MS, Alanis GA, Chaves MA, Gonzalez-Bulnes A. Effects of treatment with a prostaglandin analogue on developmental dynamics and functionality of induced corpora lutea in goats. *Anim Reprod Sci* 2010; **118**(1): 42-7.
- [28] Cognié Y, Baril G, Poulin N, Mermillod P. Current status of embryo technologies in sheep and goat. *Theriogenology* 2003; **59**(1): 171-88.
- [29] Battye KM, Fairclough RJ, Cameron AW, Trounson AO. Evidence for prostaglandin involvement in early luteal regression of the superovulated nanny goat (*Capra hircus*). *J Reprod Fertil* 1988; **84**(2): 425-30.
- [30] Cervantes MJ, Juárez ML, Mejía VO, Berruecos VJ, Vera AH, Valencia J. Use of fluorogestone acetate after breeding to reduce the effect of premature luteal regression in dairy goats when superovulation is induced with FSH. *Anim Reprod Sci* 2007; **97**(1-2): 47-54.
- [31] Holtz W, Sohnrey B, Gerland M, Driancourt MA. Ovsynch synchronization and fixed-time insemination in goats. *Theriogenology* 2008; **69**(7): 785-92.
- [32] Nur Z, Nak Y, Nak D, Üstüner B, Tuna B, Şimşek G, et al. The use of progesterone-supplemented Co-synch and Ovsynch for estrus synchronization and fixed-time insemination in nulliparous Saanen goat. *Turk J Vet Anim Sci* 2013; **37**: 183-8.
- [33] Pursley JR, Mee MO, Wiltbank MC. Synchronization of ovulation in dairy cows using PGF2 α and GnRH. *Theriogenology* 1995; **44**(7): 915-23.