

An In Vitro Investigation of the Effects of Periostin on Osteoblastic Adhesion and Proliferation on Collagen Matrices

Received: 26 October 2022, **Revised:** 24 November 2022, **Accepted:** 30 December 2022

1. Dr. Pragyan Swain

Post Graduate Student

Department of Periodontology, Chhattisgarh Dental College and Research Institute, Rajnandgaon, Chhattisgarh.

2. Dr. Ranishree Behera

Consultant Periodontist Nigam Hospital and Advanced trauma care, Balasore, Odisha,

3. Dr. Saswati Mohanty

Post Graduate Student, Department of Periodontology

Rungta College of Dental Sciences and Research, Bhilai, Chhattisgarh

4. Dr. Shanvi

Consultant Periodontist Nigam Hospital and Advanced Trauma Care, Balasore, Odisha

5. Dr. Anindita Ghosh

Post Graduate Student, Department of Periodontology

Divya Jyoti College of Dental Sciences and Research, Modinagar, Ghaziabad, U.P.

6. Dr. Bhumika J Patel

DDS, MDS(Oral Medicine and Radiology), Howard University College of Dentistry

bhumikapatel2687@gmail.com

7. Dr. Kapil Paiwal

Professor, Department of Oral & Maxillofacial Pathology, Daswani Dental College & Research Center, Kota, India

Corresponding author

Dr. Pragyan Swain

Post Graduate Student

Department of Periodontology, Chhattisgarh Dental College and Research Institute, Rajnandgaon, Chhattisgarh

Keywords

Absorbable collagen sponge, nanocrystalline hydroxyapatite collagen, osteoblast cells, periostin

Abstract

Purpose:The goal of the research was to determine how osteoblast cell adhesion, proliferation, and activity were affected when periostin was impregnated into various collagen matrices.

Materials and Methods:Onto two distinct collagen matrices, These are the steps used to cultivate and seed Saos-2 osteoblast cells: "Absorbable collagen sponges (ACS) are in Group A, absorbable collagen sponges impregnated with human periostin recombinant are in Group B, nanocrystalline hydroxyapatite collagen (NChC) is in Group C, and NChC impregnated with human periostin recombinant is in Group D. A 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide test was performed on days 2, 5, and 7 to evaluate cell viability,

adhesion, and proliferation.” Alkaline phosphatase (ALP) test was used to examine osteoblast activity in the research groups.

Results: On days 2, 5, and 7 the osteoblast adhesion in the periostin-treated absorbable collagen matrices was statistically significantly higher than that in the untreated NcHC (P=0.001). According to the results of the ALP test, the osteoblast activity in the periostin-treated ACS was higher than that in the periostin-treated NcHC.

Conclusion:

It is clear from the results of this research that Periostin plays a substantial part in regulating the cellular response of the osteoblast cells. Moreover, it has been shown that adding periostin to the ACS improves the osteoblast-like Saos-2 cells' cell survival, proliferation, and adhesion.

1. Introduction

Scientists are always on the lookout for novel biomolecules that may control the repair of periodontal disease and stimulate bone formation. During bone and tooth formation, periostin is secreted in a disulfide-linked form; this form of the 90-kDa fasciclin family member is essential. Producing this matricellular grip protein is a collaborative effort between periodontal tendon fibroblasts and periosteal osteoblasts. [2] Osteoblast grid union, mobility, grip, homeostasis, and integrity of periodontal tissues have all been shown to rely on periostin. [4,5,6] Possible therapeutic potential of the extracellular grid particle periostin, which promotes bone formation in vivo and increases the activity of osteoblasts in vitro. This provides more evidence that periostin is critical in the processes that follow tissue injury and the subsequent healing process.

The purpose of this review is to determine whether or not impregnating different collagen grids, such as an absorbable collagen wipe (ACS) or a nanocrystalline hydroxyapatite collagen (NcHC) network, with recombinant human periostin affects the growth and activity of osteoblasts. Our in vitro research hypothesized that including periostin in collagen grids would dramatically increase osteoblast activity in these networks, which was supported by our

knowledge of periostin's involvement from preclinical review models.

2. Materials And Methods

This 80% fusion was achieved by cultivating the Saos-2 osteoblast cell line, which was received from India. The four meetings of the review were as follows: Cells were cultivated on either an ACS collagen grid (Kolspon-Eucare medications, Source: Fish beginning) or a NcHC lattice (Sybograp-GBR, Eucare Pharmaceuticals).

- “Group A – ACS without any modification
- Group B – ACS impregnated with recombinant human periostin
- Group C – NcHC without any modification
- Group D – NcHC impregnated with recombinant human periostin.

Cell culture”

The cells were kept in a 37°C, 5% CO₂ humidified environment in McCoy's 5A medium containing sodium bicarbonate, 10% foetal bull like serum, L-Glutamine, and a counter-agent poison antimycotic plan. Trypsin and ethylenediaminetetraacetic acid were used to remove cells from the enhancing surface using a destructive method. Thus, cells at passages 5 and 6 were employed for this analysis, and they were suspended at a density of 10⁶ cells/ml in the culture fluid for their respective growth conditions.

Standardization of periostin for adhesion

“Human recombinant periostin (R and D Systems, Minneapolis, USA; Source: Mouse myeloma cell line) was distributed in triplicates across a sterile petri dish at doses of 5, 10, 25, 50, and 100 ng/ml.” After being rinsed with 1 phosphate-buffered saline twice and drying overnight, the plates were ready for use. The cells were trypsinized the day before and then diluted in 15 ml of McCoy basal media the next day. The resulting cell suspension was poured onto the Petri plate, and the dish was placed in an incubator with 5% CO₂ for 24 hours. When the cells were dried, they were stained with crystal violet and examined under a microscope. Cell adhesion was highest at 50 ng/ml of periostin, more than at any of the lower doses tested. Hence, 50 ng/ml was selected as the optimum concentration at which to proceed.

3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide assay

To evaluate cell viability, a 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay was used on two groups of Saos-2 cells: one treated with recombinant human periostin and the other not treated with periostin. Each social gathering was evaluated on days 2, 5, and 7. The MTT assay was also used to evaluate the viability of osteoblast cells on networks (ACS and NcHC) treated with and without recombinant periostin. Finally, the 5 mm 5 mm networks were placed in each well of a 24-well culture plate. After suffering on stage for 24 hours, 50 ng/ml of human recombinant periostin was introduced. At 37 degrees Celsius, 1 10⁴ Saos-2 cells were added to each well in a three-stage, carbon dioxide-filled configuration. The MTT test was performed on days 2, 5, and 7 of the end of the world. The review was received, and immediately a 20-liter expansion of MTT reagent (thiazolyl blue tetrazolium bromide,

5 mg/ml) was generated, and the example was tormented for 4 hours at 37°C in a CO₂ hatchery. All of the flowers and plants were thrown away, and then 100 liters of stale isopropanol were poured and allowed to bubble for an hour at 37 degrees Celsius. Absorbance measurements taken at 570 nm were saved on a microplate reader. The aforementioned technique was carried out without the cultivation of recombinant human periostin for grids that were not treated with periostin.

Alkaline phosphatase activity

Following the manufacturer's instructions, we measured basic phosphatase (High mountain) using a human High mountain protein linked immunosorbent examination pack (Bioassay Innovation Lab, China). High mountain measure was applied to cells on both periostin- and periostin-deficient grids on days 7 and 14. Measurements of 450 nm absorbance were recorded and stored in a microplate peruser. Grids that were not treated with periostin were processed by the aforementioned procedure, but without the introduction of recombinant human periostin.

Statistical analysis

The data analysis was performed using version 22 of the SPSS programming language (IBM, India). All displays of data were arranged by mean SD. Kolmogorov-Smirnov and Shapiro-Wilk tests, as well as other commonness tests, have shown that MTT absorbance is normally distributed. Hence, a t-test for independent samples was used to analyze the significance of the mean-absorbance differences. The Mann-Whitney test was employed to compare the groups since the High mountain test results did not have a normal distribution.

3. Results

On day 2, the non-periostin-treated Saos-2 cells had a higher mean examination value. Nevertheless, a higher level of MTT activity was seen in the periostin-treated wells throughout all remaining time points. “This

is indicative of a more pronounced degree of osteoblastic activation (bond and expansion) on days 5 and 7 in the periostin-treated wells. Both of these differences between the groups were very significant ($P=0.18$ and $P=0.02$, respectively) [Table 1].

Table 1

3-(4,5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide assay values of Saos-2 cells treated and not treated with periostin

Cells	MTT absorbance at 570 nm		
	2 nd day	5 th day	7 th day
Saos-2 cells	0.11±0.02	0.14±0.0095	0.221±0.02
Saos-2 cells with periostin	0.03±0.02	0.16±0.02	0.33±0.05
<i>P</i>	0.004	0.18	0.02

Periostin impregnated ACS had substantially greater osteoblast adhesion on days 5 and 7 ($P=0.013$, $P=0.001$) compared to ACS group not treated with periostin.” Similar to how periostin-treated NcHC matrices had somewhat increased osteoblast adhesion

across all time points, untreated NcHC matrices did not. ($P = 0.55$, $P = 0.87$, $P = 0.22$). Yet, there is no time point where the difference is statistically significant [Table 2].

Table 2

3-(4,5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide assay values of matrices treated and not treated with periostin

“

Matrices	MTT absorbance at 570 nm		
	2 nd day	5 th day	7 th day
ACS	0.04±0.02	0.06±0.04	0.08±0.03
ACS - Periostin	0.05±0.04	0.10±0.03	0.12±0.05
<i>P</i>	0.115	0.013	0.001
NcHC	0.011±0.06	0.022±0.04	0.03±0.05
NcHC - Periostin	0.013±0.09	0.024±0.06	0.04±0.02
<i>P</i>	0.55	0.87	0.22

By comparing ACS and NcHC, two grids that were not modified with periostin, ACS demonstrated a greater and really enormous osteoblast grip. Most notably on day 2, the thing that matters is enormous. On days 2, 5, and 7, ACS clusters had a stronger osteoblast

grip than NcHC lattice clusters, regardless of whether the grids had been treated with periostin (Collections B and D) or not. As may be observed in [Table 3], there was a significant difference between days 5 and 7 ($P=0.001$).

Table 3

3-(4,5-dimethylthiazol-2-yl)-2, 5-diphenyltetrazolium bromide assay values of Osteoblast cell line cells on matrices treated and not treated with periostin

Periostin	MTT absorbance at 570 nm	Membrane	Mean±SD	P
Cells not treated with periostin	Day 2	ACS	0.05±0.03	<0.001
		NcHC	0.03±0.03	
	Day 5	ACS	0.06±0.04	0.001
		NcHC	0.01±0.03	
Cells treated with periostin	Day 7	ACS	0.09±0.03	0.001
		NcHC	0.03±0.04	
	Day 2	ACS	0.06±0.04	0.001
		NcHC	0.01±0.04	
Day 5	ACS	0.14±0.05	<0.001	
	NcHC	0.03±0.04		
Day 7	ACS	0.2±0.04	<0.001	
	NcHC	0.02±0.07		

In the ACS and NcHC groups, Snow topped mountain activity is greater in periostin-treated grids compared to non-periostin-treated lattices on days 7 and 14, although this difference is not statistically significant.

As can be shown in Table 4, after two weeks of periostin treatment, the ACS group had more High mountain activity than the NcHC group.

Table 4

Alkaline phosphatase assay values of cells on matrices treated and not treated with periostin

Variable	Not treated with periostin	Treated with periostin	P"
ACS - 7 th day	73±6.44	112±3.44	0.2
ACS - 14 th day	156±05.34	221±7.22	0.4
P	0.04	0.02	
NcHC - 7 th day	1.9±0.44	3.89±2.22	0.25
NcHC - 14 th day	3.22±2.12	8.4±3.22	0.22
P	0.45	0.11	
ACS - 14 th day	159.3±2.11	222±6.14	
NcHC - 14 th day	3.11±2.22	7.59±3.12	
P	0.02	0.01	

4. Discussion

The alveolar bone should heal with the help of suitable signals, cells, blood flow, and supporting structures. The osteopromotive potential of today's clinical systems is

enhanced by fusing biomolecules like growth factors or proteins into their platforms. It has been shown that the bone morphogenetic proteins BMPs 2,4,7 [7,8,9], the polish grid derivative (PGD)[10], and periostin operate

as flagging atoms that accelerate the activity of designated cells, which shows promise for hastening the healing of periodontal injury. Specifically, periostin increases motility and adhesion in fibroblasts and osteoblasts found in the periodontal ligament. [11] Via the Akt/protein kinase B pathway, it controls cell adhesion and motility by binding to integrins produced by osteoblasts, such as v3 and v5. [4,5,6] Bone development in vivo and in vitro are both improved by periostin's presence. [12] The combination of these cellular processes makes periostin an important signalling molecule in bone growth and metabolism. [3] Impregnated upon ACS and nanocrystalline hydroxyapatite, recombinant human periostin was tested for its effect on osteoblastic proliferation, adhesion, and activity. Cells treated with periostin were shown to proliferate and adhere to one another more rapidly than untreated cells did over the course of many days of incubation. Results suggested that the effect of periostin on osteoblast activity followed a temporal trend. Osteoblast adhesion was shown to be significantly higher in the periostin-treated ACS groups compared to the untreated ACS groups on days 5 and 7 (P 0.029 and P 0.002, respectively). The periostin-treated group also showed improved osteoblast adhesion in all time points and NcHC matrices. This result was not statistically significant, unfortunately. The improved osteoblastic adherence seen in periostin-treated matrices is most likely due to periostin adsorption onto the surface of the biomaterials. This is widely believed to be due to periostin's ability to modulate cell-matrix interactions. Overall, our findings corroborate previous studies showing that periostin improves osteoblast cell motility and adhesion. [13,14] Comparison of cell adhesion at various times between ACS and non-periostin-treated

NcHC matrices showed that ACS consistently had better results. These findings point to the possibility that ACS helped osteoblastic cells adhere and multiply. Osteoblastic cells have been hypothesised to infiltrate and adhere to the collagen sponge's porous microenvironment. [15] Moreover, ACS offers a proper three-dimensional scaffold for tissue development and angioblast growth.

Neovascularization might be supported by the very porous ACS, which also allowed for cell in proliferation and precise cell dispersion.[16] For this reason, the ACS's role as a carrier device crucial to the maturation, specialisation, and regenerative capacity of alveolar bone. The adaptability of the ACS makes them useful for treating a wide variety of tissue problems, including periodontal bone deficiencies, cyst cavities, and alveolar bone augmentation. [17]

Matrix impregnated with periostin showed significantly greater osteoblast adhesion in ACS treated with periostin at all time points compared to NcHC treated with periostin. Recombinant human periostin added to ACS had a synergistic impact on osteoblastic cell behaviour, increasing both proliferation and adhesion. Because of its ability to boost cellular activity, ACS has become the standard bearer for transporting growth factors during periodontal regeneration. In a clinical setting, the application of rhBMP-2 in ACS for the correction of supra-alveolar periodontal bone deformities [18], three-walled intrabony periodontal defects [19], and edging up showed clinically considerable alveolar bone healing. [20] Although ACS is widely accepted as the gold standard for transporting natural specialists and has been clinically validated for a wide variety of uses, it is not without its drawbacks. Larger non-bone-supported deficiencies are outside the scope of ACS since the technology lacks

extended primary security, which is necessary in these circumstances. Examples of such deficiencies are those with just one wall or locations with vertical extension destinations.

Overall, however, we did not find any statistically significant differences between ACS groups treated with periostin and those that weren't, and the high mountain action in the periostin-treated group was more pronounced than that in the control group. Compared to the NcHC group, the ACS group's snowy mountain activity increased in the first and second prolonged times after receiving periostin. Consider the crucial capacity provided by the ACS architecture in regulating osteoblast grip, and the increased high-mountain activity may make more sense.

The discovery that periostin may coordinate the cell response on these collagen grids has led to significant progress in the area of periodontal bone repair. While most of the matrices utilised in this investigation were collagen-based, they varied in terms of their micro- and macrostructure. Bioactivity of periostin impregnated into NcHC matrices was reduced, likely because nanohydroxyapatite impregnated into collagen rendered the NcHC matrix resistant to the periostin. Impregnating matrices with periostin seems to be a promising strategy for controlling host osteoblast activity. The next step in the trial procedure is to evaluate the effect of periostin in well-designed, longitudinally-monitored animal experiments.

5. Conclusion

This in vitro analysis lends credence to the idea that osteoblast-like Saos-2 cells' survival, proliferation, and adhesion may be influenced by impregnating ACS with recombinant human periostin. The

periodontal benefits of periostin may be addressed by a naturally occurring expert with a high degree of intelligence. Yet, periodontal applications need an in-depth understanding of periostin's bioactivity in the periodontium.

References

- [1] Horiuchi K, Amizuka N, Takeshita S, Takamatsu H, Katsuura M, Ozawa H, et al. Identification and characterization of a novel protein, periostin, with restricted expression to periosteum and periodontal ligament and increased expression by transforming growth factor beta. *J Bone Miner Res.* 1999;14:1239–49.
- [2] Suzuki H, Amizuka N, Kii I, Kawano Y, Nozawa-Inoue K, Suzuki A, et al. Immunohistochemical localization of periostin in tooth and its surrounding tissues in mouse mandibles during development. *Anat Rec A Discov Mol Cell Evol Biol.* 2004;281:1264–75.
- [3] Merle B, Garnero P. The multiple facets of periostin in bone metabolism. *Osteoporos Int.* 2012;23:1199–212.
- [4] Shimazaki M, Nakamura K, Kii I, Kashima T, Amizuka N, Li M, et al. Periostin is essential for cardiac healing after acute myocardial infarction. *J Exp Med.* 2008;205:295–303.
- [5] Ouyang G, Liu M, Ruan K, Song G, Mao Y, Bao S. Upregulated expression of periostin by hypoxia in non-small-cell lung cancer cells promotes cell survival via the Akt/PKB Pathway. *Cancer Let.* 2009;281:213–9.
- [6] Kühn B, del Monte F, Hajjar RJ, Chang YS, Lebeche D, Arab S, et al. Periostin induces proliferation of differentiated cardiomyocytes and promotes cardiac repair. *Nat Med.* 2007;13:962–9.

- [7] Zhao M, Xiao G, Berry JE, Franceschi RT, Reddi A, Somerman MJ. Bone morphogenetic protein 2 induces dental follicle cells to differentiate toward a cementoblast/osteoblast phenotype. *J Bone Miner Res.* 2002;17:1441–51.
- [8] Ahn SH, Kim CS, Suk HJ, Lee YJ, Choi SH, Chai JK, et al. Effect of recombinant human bone morphogenetic protein-4 with carriers in rat calvarial defects. *J Periodontol.* 2003;74:787–97.
- [9] Jin QM, Zhao M, Economides AN, Somerman MJ, Giannobile WV. Noggin gene delivery inhibits cementoblast-induced mineralization. *Connect Tissue Res.* 2004;45:50–9.
- [10] Lyngstadaas SP, Lundberg E, Ekdahl H, Andersson C, Gestrelus S. Autocrine growth factors in human periodontal ligament cells cultured on enamel matrix derivative. *J Clin Periodontol.* 2001;28:181–8.
- [11] Padiol-Molina M, Volk SL, Rios HF. Periostin increases migration and proliferation of human periodontal ligament fibroblasts challenged by tumor necrosis factor α and *Porphyromonas gingivalis* lipopolysaccharides. *J Periodontol Res.* 2014;49:405–14.
- [12] Heo SC, Shin WC, Lee MJ, Kim BR, Jang IH, Choi EJ, et al. Periostin accelerates bone healing mediated by human mesenchymal stem cell-embedded hydroxyapatite/tricalcium phosphate scaffold. *PLoS One.* 2015;10 doi:10.1371/journal.pone.0116698:1-17. 13. Zhu S, Barbe MF, Liu C, Hadjiargyrou M, Popoff SN, Rani S, et al. Periostin-like-factor in osteogenesis. *J Cell Physiol.* 2009;218:584–92.
- [13] Rani S, Barbe MF, Barr AE, Litivn J. Role of TNF alpha and PLF in bone remodeling in a rat model of repetitive reaching and grasping. *J Cell Physiol.* 2010;225:152–67.
- [14] Wang HL, Miyauchi M, Takata T. Initial attachment of osteoblasts to various guided bone regeneration membranes: An *in vitro* study. *J Periodontal Res.* 2002;37:340–4.
- [15] Geiger M, Li RH, Friess W. Collagen sponges for bone regeneration with rhBMP-2. *Adv Drug Deliv Rev.* 2003;55:1613–29.
- [16] Matsuno T, Nakamura T, Kuremoto K, Notazawa S, Nakahara T, Hashimoto Y, et al. Development of beta-tricalcium phosphate/collagen sponge composite for bone regeneration. *Dent Mater J.* 2006;25:138–44.
- [17] Sigurdsson TJ, Nguyen S, Wikesjo UM. Alveolar ridge augmentation with rhBMP-2 and bone-to-implant contact in induced bone. *Int J Periodontics Restorative Dent.* 2001;21:461–73.
- [18] Chi SH, Kim CK, Cho KS, Huh JS, Sorensen RG, Wozney JM, et al. Effect of recombinant human morphogenetic protein-2/Absorbable Collagen Sponge on healing in 3-wall intrabony defect in dogs. *J periodontol.* 2002;73:63–72.
- [19] Barboza EP, Caula AL, de Oliveria Caula F, de Souza RO, Neto LG, Sorensen RG, et al. Effect of recombinant human morphogenetic protein-2 in an Absorbable Collagen Sponge with space-providing biomaterials on the augmentation of chronic alveolar ridge defect. *J Periodontol.* 2004;75:702–8.